

ERYTHROCYTE MEASUREMENTS IN BIRDS

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GULLIVER (1840, 1846, 1875) appears to have been the earliest to make an extensive study of the size of red cells in vertebrates, including birds. Cleland and Johnston (1912) measured the erythrocytes in 90 species of birds distributed among 30 families. Bartsch, Ball, Rosenzweig, and Salman (1937) reported measurements in 50 North American birds. However, only two of their species were studied by us. Lucas and Jamroz (1961) described the various blood cell types in the fowl but they give no cell measurements for species other than *Gallus domesticus*.

Measurements of red cell size give some idea of the surface area offered for exchange of gases with the plasma. The least diameter of elliptical erythrocytes is a gauge of minimal capillary diameter for a particular species. Erythrocyte size appears to be related to the general metabolic activity of the species. For these reasons such information is valuable in a comparative study.

While collecting specimens for adrenal and other studies we were frequently presented with the opportunity to make fresh blood smears from the animals. Material so obtained was used as a basis for this study. Our tropical material came from Panamá while the remainder was from the United States, mostly Ohio and Florida.

METHODS

Blood smears were made immediately after the death of the animals. These were dried promptly and stored until prepared for study in the laboratory. Staining was carried out under controlled conditions. Five drops of standard Wright's stain were allowed to remain on the slide for one minute before addition of a pH 6.5 buffer. The slide was allowed to stand for five minutes at room temperature and then washed with distilled water for 30 seconds and allowed to dry.

Ten cells on each slide, selected for excellence of staining and internal cytology, were measured by means of a calibrated eyepiece used in conjunction with an oil-immersion objective. In this way 10 measurements of maximum cell width and length as well as nuclear width and length were made. The ratios of cytosome length to width and nucleus length to width have been calculated. This is a measure of cell and nuclear deviation from a spherical shape. The surface area of the cell is an important parameter, but was not calculated because cell thickness in these air-dried specimens could not be determined.

RESULTS

In this survey, 124 species distributed among 46 families are included (Table 1). When measurements are of only one individual of a species, the standard error of the values for the different cells is shown, while when three or more individual specimens of a species are measured the standard error is based on the averages of the individuals involved.

TABLE 1
ERYTHROCYTE MEASUREMENTS IN BIRDS

Family and species	Cytosome			Nucleus		
	Length (μ)	Width (μ)	Ratio L/W	Length (μ)	Width (μ)	Ratio L/W
Tinamidae						
<i>Crypturellus soui</i>	13.5 \pm 0.24	7.6 \pm 0.26	1.87	5.2 \pm 0.19	2.3 \pm 0.11	2.26
" "	13.9 \pm 0.19	7.5 \pm 0.12	1.85	4.4 \pm 0.10	2.5 \pm 0.06	1.76
Podicipedidae						
<i>Podiceps dominicus</i>	12.7 \pm 0.80	8.2 \pm 0.80	1.60	5.1 \pm 0.50		
" "	13.7 \pm 0.80	8.0 \pm 0.30	1.71	5.7 \pm 0.80	2.8 \pm 0.30	2.03
<i>Podilymbus podiceps</i>	14.6 \pm 0.26	8.2 \pm 0.11	1.87	5.2 \pm 0.12	2.2 \pm 0.02	2.36
Pelecanidae						
<i>Pelecanus occidentalis</i> (10)	14.2 \pm 0.20	7.8 \pm 0.09	1.75	5.9 \pm 0.12	2.9 \pm 0.065	2.03
Phalacrocoracidae						
<i>Phalacrocorax auritus</i> (6)	13.6 \pm 0.17	7.6 \pm 0.17	1.75	5.2 \pm 0.11	2.6 \pm 0.08	2.00
Anhingidae						
<i>Anhinga anhinga</i>	15.1 \pm 0.52	8.6 \pm 0.30	1.54	6.5 \pm 0.37	3.0 \pm 0.30	2.17
" "	15.7 \pm 0.19	10.5 \pm 0.33	1.57	5.4 \pm 0.19		
Ardeidae						
<i>Ardea herodias</i>	14.3 \pm 0.92	7.4 \pm 0.68	2.00	5.8 \pm 0.56	3.0 \pm 0.34	1.93
<i>Butorides virescens</i>	13.0 \pm 0.22	8.3 \pm 0.15	1.62	6.0 \pm 0.16	2.9 \pm 0.17	2.07
<i>Florida caerulea</i>	12.8 \pm 0.64	7.3 \pm 0.49	1.85	6.1 \pm 0.33	3.0 \pm 0.15	2.03
<i>Casmerodius albus</i> (3)	15.0 \pm 0.39	7.9 \pm 0.30	1.87	6.7 \pm 0.33	3.3 \pm 0.23	2.03
<i>Tigrisoma lineatum</i>	15.8 \pm 0.37	10.2 \pm 0.17	1.58	7.0 \pm 0.22	2.9 \pm 0.19	2.41
<i>Leucophox thula</i>	13.1 \pm 0.38	7.7 \pm 0.16	1.70	5.2 \pm 0.12	2.6 \pm 0.03	2.00
Cathartidae						
<i>Sarcorampus papa</i>	14.6 \pm 0.34	8.5 \pm 0.40	1.79	6.7 \pm 0.34	3.3 \pm 0.14	2.03
" "	14.0 \pm 0.87	7.6 \pm 0.43	1.84	6.2 \pm 0.33	3.0 \pm 0.17	2.06
<i>Coragyps atratus</i> (5)	14.0 \pm 0.20	7.7 \pm 0.11	1.81	6.3 \pm 0.20	2.6 \pm 0.17	2.42
<i>Cathartes aura</i> (3)	14.0 \pm 0.32	7.5 \pm 0.13	1.87	6.0 \pm 0.18	2.4 \pm 0.07	2.50
Accipitridae						
<i>Accipiter cooperii</i>	14.3 \pm 0.27	8.1 \pm 0.13	1.77	6.2 \pm 0.14	2.4 \pm 0.11	2.58
<i>Buteo platypterus</i>	13.4 \pm 0.51	7.6 \pm 0.34	1.77	6.2 \pm 0.31	3.0 \pm 0.32	2.07
" "	13.7 \pm 0.46	7.8 \pm 0.13	1.76	5.5 \pm 0.21	2.4 \pm 0.17	2.29
Falconidae						
<i>Falco sparverius</i>	11.8 \pm 0.18	7.3 \pm 0.17	1.62	5.8 \pm 0.13	2.1 \pm 0.04	2.76
<i>Caracara cheriway</i>	14.1 \pm 0.05	8.7 \pm 0.16	1.62	6.4 \pm 0.13	2.7 \pm 0.11	2.37
Cracidae						
<i>Chamaepetes unicolor</i>	13.6 \pm 0.52	7.6 \pm 0.42	1.84	6.1 \pm 0.53	2.7 \pm 0.22	2.26
" "	14.1 \pm 0.75	7.4 \pm 0.35	1.91	6.9 \pm 0.65	2.9 \pm 0.21	2.38
Phasianidae						
<i>Odontophorus guttatus</i>	11.9 \pm 0.46	6.8 \pm 0.28	1.75	5.2 \pm 0.41	2.3 \pm 0.21	2.26
Rallidae						
<i>Rallus elegans</i>	14.5 \pm 0.32	7.7 \pm 0.17	1.89	5.7 \pm 0.13	2.9 \pm 0.12	1.97
<i>Aramides cajanea</i>	12.9 \pm 0.78	7.2 \pm 0.50	1.79	5.5 \pm 0.40	3.3 \pm 0.37	1.67
<i>Laterallus albigularis</i>	12.3 \pm 0.60	7.3 \pm 0.39	1.68	5.3 \pm 0.43	3.2 \pm 0.20	1.65
" "	13.2 \pm 0.21	7.1 \pm 0.10	1.86	5.1 \pm 0.09	3.0 \pm 0.07	1.70
<i>Porphyryla martinica</i>	14.2 \pm 0.49	7.8 \pm 0.24	1.82	6.7 \pm 0.53	2.9 \pm 0.21	2.31
<i>Fulica americana</i>	11.4 \pm 0.26	7.5 \pm 0.22	1.51	4.2 \pm 0.06	2.3 \pm 0.07	1.83
Heliornithidae						
<i>Heliornis julica</i>	12.6 \pm 0.15	7.5 \pm 0.14	1.68	5.5 \pm 0.21	3.0 \pm 0.11	1.83
Jacaniidae						
<i>Jacana spinosa</i>	13.7 \pm 0.79	7.5 \pm 0.52	1.83	5.9 \pm 0.64	3.7 \pm 0.33	1.59
Charadriidae						
<i>Charadrius wilsonia</i> (6)	12.8 \pm 0.27	7.3 \pm 0.13	1.76	5.8 \pm 0.11	2.4 \pm 0.05	2.41
Recurvirostridae						
<i>Himantopus mexicanus</i>	12.8 \pm 0.16	6.9 \pm 0.18	1.85	5.8 \pm 0.20	2.5 \pm 0.99	2.32
Columbidae						
<i>Columba albilinea</i>	12.1 \pm 0.56	7.7 \pm 0.29	1.67	5.8 \pm 0.39	2.7 \pm 0.31	2.15
" "	13.4 \pm 0.67	7.5 \pm 0.21	1.78	6.4 \pm 0.13	2.7 \pm 0.12	2.37
<i>Columbigallina minuta</i>	12.3 \pm 0.87	7.4 \pm 0.40	1.66	5.9 \pm 0.43	2.5 \pm 0.13	2.36
<i>Columbigallina talpacoti</i>	11.6 \pm 0.28	7.0 \pm 0.30	1.66	6.4 \pm 0.31	2.4 \pm 0.27	2.67
<i>Geotrygon chiriquensis</i>	12.0 \pm 0.59	6.9 \pm 0.32	1.72	5.6 \pm 0.38	2.8 \pm 0.28	2.00
" "	13.2 \pm 0.48	7.7 \pm 0.43	1.71	6.4 \pm 0.38	3.3 \pm 0.24	1.94
Psittacidae						
<i>Pyrrhura hoffmanni</i>	13.2 \pm 0.23	7.2 \pm 0.18	1.83	6.1 \pm 0.06	2.3 \pm 0.04	2.65
<i>Brologeris jugularis</i>	12.1 \pm 0.57	7.7 \pm 0.18	1.75	6.0 \pm 0.32	2.9 \pm 0.25	2.07
<i>Pionus senilis</i>	13.0 \pm 0.47	7.6 \pm 0.25	1.72	5.8 \pm 0.32	2.3 \pm 0.04	2.52
<i>Amazona autumnalis</i>	13.6 \pm 0.74	7.7 \pm 0.01	1.77	6.2 \pm 0.54	3.0 \pm 0.56	2.07

TABLE 1—Continued

Family and species	Cytosome			Nucleus		
	Length (μ)	Width (μ)	Ratio L/W	Length (μ)	Width (μ)	Ratio L/W
Cuculidae						
<i>Piaya cayana</i>	13.7 \pm 0.19	7.9 \pm 0.13	1.73	6.2 \pm 0.13	2.9 \pm 0.09	2.14
<i>Crotophaga major</i>	12.0 \pm 0.64	8.1 \pm 0.84	1.48	5.5 \pm 0.35	3.4 \pm 0.34	1.62
<i>Crotophaga ani</i> (4)	12.9 \pm 0.22	6.9 \pm 0.26	1.87	5.8 \pm 0.28	2.9 \pm 0.29	2.00
Strigidae						
<i>Otus choliba</i>	12.6 \pm 0.96	7.5 \pm 0.33	1.68	6.0 \pm 0.33	3.1 \pm 0.12	1.94
" "	13.9 \pm 0.35	7.4 \pm 0.15	1.88	5.4 \pm 0.25	2.9 \pm 0.03	1.86
<i>Strix varia</i>	13.8 \pm 0.24	7.3 \pm 0.23	1.82	5.6 \pm 0.20	2.7 \pm 0.14	2.07
" "	13.5 \pm 0.28	7.8 \pm 0.16	1.73	5.7 \pm 0.11	2.2 \pm 0.04	2.59
<i>Rhinophlynx clamator</i> (4)	13.7 \pm 0.22	7.7 \pm 0.29	1.80	5.9 \pm 0.18	3.0 \pm 0.21	1.96
<i>Speotyto cunicularia</i>	14.1 \pm 0.11	7.8 \pm 0.25	1.81	6.3 \pm 0.32	2.7 \pm 0.12	2.33
Nyctibiidae						
<i>Nyctibius griseus</i>	14.4 \pm 0.77	8.2 \pm 0.52	1.77	6.3 \pm 0.30	3.4 \pm 0.34	1.85
" "	13.2 \pm 0.56	7.7 \pm 0.47	1.71	6.4 \pm 0.31	3.3 \pm 0.22	1.94
Caprimulgidae						
<i>Nyctidromus albicollis</i> (3)	13.6 \pm 0.53	8.3 \pm 0.56	1.64	6.3 \pm 0.23	3.3 \pm 0.80	1.91
Trochilidae						
<i>Glaucis hirsuta</i>	12.2 \pm 0.43	6.9 \pm 0.37	1.77	6.2 \pm 0.37	2.8 \pm 0.20	2.21
<i>Phaethornis guy</i>	11.5 \pm 0.26	6.3 \pm 0.10	1.77	4.5 \pm 0.23	2.1 \pm 0.04	2.14
" "	12.0 \pm 0.14	7.0 \pm 0.20	1.71	5.2 \pm 0.19	2.1 \pm 0.04	2.47
<i>Campylopterus hemileucurus</i>	11.1 \pm 0.37	5.6 \pm 0.35	1.98	5.0 \pm 0.19	1.8 \pm 0.25	2.77
<i>Anthracoceros nigricollis</i>	10.7 \pm 0.12	6.4 \pm 0.13	1.67	5.2 \pm 0.10	2.6 \pm 0.02	2.00
<i>Thalurania jurcata</i>	11.3 \pm 0.34	6.4 \pm 0.21	1.77	5.5 \pm 0.41	2.1 \pm 0.13	2.68
<i>Damophila julie</i>	10.9 \pm 0.38	6.1 \pm 0.36	1.79	5.2 \pm 0.17	2.6 \pm 0.10	2.00
<i>Amazilia edward</i>	11.4 \pm 0.21	6.0 \pm 0.02	1.90	5.7 \pm 0.08	2.0 \pm 0.04	2.85
<i>Amazilia tzacatl</i>	11.5 \pm 0.23	6.3 \pm 0.13	1.85	5.6 \pm 0.21	2.6 \pm 0.22	2.15
" "	10.9 \pm 0.11	6.5 \pm 0.21	1.68	5.7 \pm 0.08	2.5 \pm 0.03	2.28
<i>Heliodoxa jacula</i>	12.3 \pm 0.17	6.3 \pm 0.12	1.95	6.0 \pm 0.05	2.1 \pm 0.05	2.85
<i>Selasphorus scintilla</i>	10.7 \pm 0.85	6.1 \pm 0.46	1.75	5.7 \pm 0.26	2.5 \pm 0.11	2.28
Trogonidae						
<i>Pharomachus mocino</i>	12.2 \pm 0.57	7.0 \pm 0.47	1.74	5.8 \pm 0.34	2.7 \pm 0.17	2.15
<i>Trogon massena</i> (3)	13.6 \pm 0.39	7.6 \pm 0.05	1.79	6.3 \pm 0.14	2.6 \pm 0.35	2.42
<i>Trogon collaris</i>	13.8 \pm 0.97	7.8 \pm 0.25	1.76	7.0 \pm 0.53	2.7 \pm 0.19	2.59
" "	13.3 \pm 0.87	7.6 \pm 0.54	1.76	6.6 \pm 0.30	3.0 \pm 0.42	2.20
Alcedinidae						
<i>Chloroceryle amazona</i>	14.0 \pm 0.47	7.6 \pm 0.26	1.79	6.8 \pm 0.36	3.1 \pm 0.11	2.19
" "	13.8 \pm 0.27	7.9 \pm 0.29	1.74	6.1 \pm 0.21	2.7 \pm 0.21	2.26
<i>Chloroceryle aenea</i>	12.6 \pm 0.56	6.7 \pm 0.31	1.88	6.5 \pm 0.50	2.9 \pm 0.26	2.24
Bucconidae						
<i>Notharchus macrorhynchos</i>	13.1 \pm 0.13	7.6 \pm 0.08	1.72	6.2 \pm 0.10	2.9 \pm 0.08	2.17
Ramphastidae						
<i>Aulacorhynchus prasinus</i> (3)	12.8 \pm 0.65	8.0 \pm 0.12	1.60	7.2 \pm 0.18	2.9 \pm 0.18	2.48
<i>Pteroglossus torquatus</i> (5)	13.9 \pm 0.25	8.0 \pm 0.15	1.73	6.6 \pm 0.20	2.7 \pm 0.17	2.44
<i>Ramphastos swainsoni</i>	13.3 \pm 0.28	7.7 \pm 0.35	1.73	5.8 \pm 0.30	2.3 \pm 0.20	2.52
Picidae						
<i>Dryocopus lineatus</i>	14.3 \pm 0.55	7.6 \pm 0.23	1.88	6.4 \pm 0.39	3.1 \pm 0.24	2.06
<i>Melanerpes formicivorus</i>	12.9 \pm 0.73	7.1 \pm 0.48	1.89	6.4 \pm 0.28	2.8 \pm 0.15	2.28
" "	11.9 \pm 0.89	6.0 \pm 0.44	1.98	6.4 \pm 0.34	2.8 \pm 0.35	2.28
<i>Centurus rubricapillus</i> (6)	13.7 \pm 0.37	7.3 \pm 0.15	1.88	6.1 \pm 0.15	2.4 \pm 0.10	2.54
<i>Centurus chrysachen</i>	11.7 \pm 0.11	6.8 \pm 0.29	1.80	5.0 \pm 0.17	2.1 \pm 0.04	2.38
" "	13.0 \pm 0.19	7.0 \pm 0.16	1.84			
<i>Phloeocastes guatemalensis</i> (3)	13.8 \pm 0.29	7.15 \pm 0.33	1.84	6.0 \pm 0.29	2.5 \pm 0.27	2.40
Dendrocolaptidae						
<i>Dendrocincla homochroa</i>	11.8 \pm 0.49	6.7 \pm 0.36	1.76	5.4 \pm 0.32	2.3 \pm 0.29	2.35
<i>Sittasomus griseicapillus</i>	12.0 \pm 0.15	6.4 \pm 0.12	1.87	5.4 \pm 0.13	2.0 \pm 0.02	2.70
<i>Xiphorhynchus guttatus</i>	12.0 \pm 0.61	7.1 \pm 0.41	1.70	5.6 \pm 0.29	2.7 \pm 0.13	2.07
" "	12.5 \pm 0.62	7.4 \pm 0.43	1.69	6.4 \pm 0.50	2.8 \pm 0.20	2.29
<i>Xiphorhynchus erythropygius</i>	12.2 \pm 0.52	6.4 \pm 0.36	1.91	5.7 \pm 0.44	2.4 \pm 0.27	2.37
<i>Lepidocolaptes affinis</i>	11.2 \pm 0.80	6.6 \pm 0.55	1.70	6.0 \pm 0.34	2.6 \pm 0.13	2.31
Furnariidae						
<i>Synallaxis brachyura</i>	11.1 \pm 0.49	6.1 \pm 0.39	1.82	5.6 \pm 0.42	2.1 \pm 0.21	2.67
<i>Anabacerthia striaticollis</i>	11.9 \pm 0.17	6.1 \pm 0.01	1.95	5.4 \pm 0.15	2.1 \pm 0.05	2.57
Fornicariidae						
<i>Taraba major</i>	13.5 \pm 0.33	7.5 \pm 0.30	1.80	6.2 \pm 0.38	2.7 \pm 0.08	2.30
" "	13.7 \pm 0.67	7.6 \pm 0.26	1.80	7.0 \pm 0.55	2.9 \pm 0.25	2.41
Pipridae						
<i>Corapipo leucorrhoea</i>	11.1 \pm 0.16	6.1 \pm 0.10	1.82	5.1 \pm 0.11	2.3 \pm 0.07	2.22
<i>Manacus vitellinus</i>	12.6 \pm 0.26	7.1 \pm 0.58	1.67	6.2 \pm 0.48	3.0 \pm 0.30	2.07
" "	11.3 \pm 0.29	6.2 \pm 0.42	1.82	5.6 \pm 0.26	2.7 \pm 0.20	2.07

TABLE 1—Continued

Family and species	Cytosome			Nucleus		
	Length (μ)	Width (μ)	Ratio L/W	Length (μ)	Width (μ)	Ratio L/W
Cotingidae						
<i>Cotinga ridgwayi</i>	12.2 \pm 0.27	6.6 \pm 0.30	1.80	6.2 \pm 0.21	2.2 \pm 0.19	2.82
" "	12.1 \pm 0.26	6.8 \pm 0.08	1.78	6.2 \pm 0.19	2.0 \pm 0.05	3.10
<i>Attila spadiceus</i>	12.3 \pm 0.67	6.6 \pm 0.42	1.86	5.7 \pm 0.30	1.9 \pm 0.24	3.00
<i>Tityra semifasciata</i>	11.7 \pm 0.63	6.8 \pm 0.38	1.72	5.6 \pm 0.40	2.7 \pm 0.22	2.17
Tyrannidae						
<i>Myiozetetes cayenensis</i>	10.7 \pm 0.12	6.3 \pm 0.10	1.81	5.2 \pm 0.82	2.5 \pm 0.06	2.08
<i>Contopus virens</i> (3)	11.9 \pm 0.11	6.8 \pm 0.05	1.75	6.0 \pm 0.30	2.5 \pm 0.20	2.40
<i>Empidonax flaviventris</i>	12.2 \pm 0.50	5.6 \pm 0.29	2.06	6.0 \pm 0.33	2.1 \pm 0.30	2.86
" "	11.8 \pm 0.46	6.2 \pm 0.37	1.90	5.4 \pm 0.25	2.1 \pm 0.13	2.57
<i>Mitrephanes phaeocercus</i>	11.1 \pm 0.44	6.4 \pm 0.28	1.73	5.6 \pm 0.35	2.1 \pm 0.13	2.67
<i>Todirostrum cinereum</i>	11.9 \pm 0.90	6.4 \pm 0.29	1.86	5.7 \pm 0.47	2.3 \pm 0.21	2.48
<i>Lophotriccus pileatus</i>	11.5 \pm 0.14	6.4 \pm 0.20	1.79	4.7 \pm 0.10	2.2 \pm 0.04	2.14
Hirundinidae						
<i>Progne chalybea</i> (3)	12.7 \pm 0.70	7.1 \pm 0.08	1.79	5.5 \pm 0.38	2.7 \pm 0.21	2.04
<i>Stelgidopteryx ruficollis</i> (3)	12.2 \pm 0.27	6.6 \pm 0.25	1.90	5.9 \pm 0.11	2.1 \pm 0.09	2.81
<i>Pygocelidon cyanoleuca</i> (3)	11.7 \pm 0.19	6.4 \pm 0.16	1.80	5.4 \pm 0.15	2.1 \pm 0.09	2.57
Corvidae						
<i>Cyanocitta cristata</i>	13.7 \pm 0.14	7.8 \pm 0.92	1.77	5.8 \pm 0.12	2.6 \pm 0.06	2.23
Troglodytidae						
<i>Thryothorus modestus</i>	10.8 \pm 0.20	6.5 \pm 0.07	1.66	5.6 \pm 0.10	2.6 \pm 0.02	2.15
<i>Thryothorus fascioventris</i>	12.1 \pm 0.56	6.9 \pm 0.48	1.75	5.5 \pm 0.33	2.9 \pm 0.23	1.90
<i>Troglodytes musculus</i>	10.9 \pm 0.23	6.7 \pm 0.22	1.72	5.5 \pm 0.28	2.2 \pm 0.10	2.50
" "	11.3 \pm 0.22	6.2 \pm 0.17	1.82	5.2 \pm 0.17	2.1 \pm 0.06	2.48
Turdidae						
<i>Turdus grayi</i>	12.1 \pm 0.08	6.7 \pm 0.15	1.81	5.4 \pm 0.11	2.0 \pm 0.06	2.70
<i>Turdus plebejus</i>	11.8 \pm 0.60	7.3 \pm 0.46	1.62	5.5 \pm 0.29	2.8 \pm 0.25	1.97
Bombycillidae						
<i>Bombycilla cedrorum</i> (4)	12.2 \pm 0.26	6.3 \pm 0.13	1.93	5.7 \pm 0.22	2.3 \pm 0.07	2.48
Cyclarhidae						
<i>Cyclarhis gujanensis</i>	11.8 \pm 0.71	6.6 \pm 0.67	1.79	5.9 \pm 0.20	2.6 \pm 0.19	2.27
"Coerebidae"						
<i>Dacnis cayana</i>	12.5 \pm 0.43	7.4 \pm 0.31	1.69	6.2 \pm 0.26	3.2 \pm 0.19	1.94
<i>Dacnis venusta</i>	11.2 \pm 0.18	6.0 \pm 0.06	1.87	4.8 \pm 0.13	2.1 \pm 0.04	2.29
Parulidae						
<i>Mniotilta varia</i>	11.2 \pm 0.17	6.1 \pm 0.10	1.84	4.9 \pm 0.16	2.3 \pm 0.05	2.13
<i>Vermivora gutturalis</i>	11.1 \pm 0.50	7.2 \pm 0.52	1.82	5.6 \pm 0.42	2.9 \pm 0.25	1.93
<i>Dendroica pennsylvanica</i>	11.7 \pm 0.34	6.5 \pm 0.32	1.80	5.9 \pm 0.24	2.2 \pm 0.11	2.68
<i>Geothlypis chiriquirensis</i>	11.9 \pm 0.18	5.9 \pm 0.06	2.02	5.3 \pm 0.16	2.0 \pm 0.03	2.65
<i>Wilsonia pusilla</i>	11.6 \pm 0.40	5.9 \pm 0.25	2.06	5.7 \pm 0.40	2.5 \pm 0.24	2.28
" "	11.8 \pm 0.14	5.6 \pm 0.15	2.11	5.5 \pm 0.12	2.0 \pm 0.03	2.75
<i>Myioborus miniatus</i>	11.6 \pm 0.15	6.1 \pm 0.07	1.90	5.1 \pm 0.14	2.3 \pm 0.07	2.22
<i>Basileuterus rufifrons</i>	11.5 \pm 0.33	6.0 \pm 0.36	1.91	5.6 \pm 0.28	2.6 \pm 0.66	2.15
Icteridae						
<i>Zarkynchus wagleri</i>	10.6 \pm 0.39	7.1 \pm 0.19	1.65	5.0 \pm 0.12	2.1 \pm 0.05	2.38
" "	11.4 \pm 0.24	6.2 \pm 0.21	1.84	4.8 \pm 0.15	2.2 \pm 0.07	2.18
<i>Amblycercus holosericeus</i>	11.6 \pm 0.57	6.5 \pm 0.27	1.77	5.7 \pm 0.47	3.0 \pm 0.23	1.90
<i>Icterus mesomelas</i>	13.1 \pm 0.54	6.9 \pm 0.45	1.90	5.7 \pm 0.36	2.8 \pm 0.20	2.04
<i>Icterus galbula</i> (3)	12.0 \pm 0.30	6.3 \pm 0.20	1.90	5.3 \pm 0.21	2.2 \pm 0.07	2.41
Thraupidae						
<i>Tanagra icterocephala</i>	11.4 \pm 0.07	6.3 \pm 0.21	1.82	5.1 \pm 0.19	2.4 \pm 0.18	2.13
<i>Thraupis episcopus</i>	11.7 \pm 0.33	7.0 \pm 0.19	1.62	5.7 \pm 0.38	2.3 \pm 0.21	2.48
" "	11.0 \pm 0.27	7.0 \pm 0.09	1.57	5.0 \pm 0.15	2.2 \pm 0.02	2.27
<i>Ramphocelus passerinii</i>	12.0 \pm 0.18	6.6 \pm 0.13	1.82	4.8 \pm 0.08	2.4 \pm 0.04	2.00
<i>Chlorospingus pileatus</i>	12.0 \pm 0.86	6.7 \pm 0.50	1.71	5.9 \pm 0.25	2.6 \pm 0.00	2.27
" "	10.8 \pm 0.57	6.6 \pm 0.46	1.64	5.8 \pm 0.39	2.7 \pm 0.20	2.15
Fringillidae						
<i>Saltator atriceps</i>	12.5 \pm 0.49	7.5 \pm 0.64	1.66	5.3 \pm 0.22	2.9 \pm 0.20	1.83
<i>Saltator albicollis</i>	12.2 \pm 0.68	7.6 \pm 0.28	1.68	5.2 \pm 0.42	3.1 \pm 0.19	1.68
" "	12.0 \pm 0.18	6.8 \pm 0.15	1.77	5.2 \pm 0.12	2.8 \pm 0.09	1.86
<i>Pheucticus tibialis</i>	11.7 \pm 0.63	6.5 \pm 0.49	1.80	5.3 \pm 0.28	2.7 \pm 0.23	1.92
<i>Tiaris olivacea</i>	11.3 \pm 0.20	6.9 \pm 0.25	1.64	4.9 \pm 0.14	2.4 \pm 0.08	2.04
<i>Arremonops conirostris</i>	11.0 \pm 0.21	6.2 \pm 0.04	1.83	5.5 \pm 0.15	2.2 \pm 0.05	2.50
" "	11.7 \pm 0.15	6.2 \pm 0.08	1.88	5.4 \pm 0.15	2.1 \pm 0.04	2.57
<i>Zonotrichia capensis</i> (5)	11.4 \pm 0.15	6.1 \pm 0.15	1.88	4.8 \pm 0.07	2.1 \pm 0.06	2.29

The range of measurements of the erythrocytes is from $10.7 \mu \times 6.1 \mu$, in *Selasphorus*, to $15.8 \mu \times 10.2 \mu$, in *Tigrisoma*. The larger erythrocytes appear in the lower forms. Most of the passerines and trochilids contain the smaller red cells. The corvid *Cyanocitta* contains large erythrocytes ($13.7 \mu \times 7.8 \mu$). In a few families these cells are smaller in the smaller species. This is so in the ardeids (*Butorides*, $13.0 \mu \times 8.3 \mu$; *Florida*, $12.8 \mu \times 7.3 \mu$; *Leucophoyx*, $13.1 \mu \times 7.7 \mu$ compared with *Ardea*, $14.3 \mu \times 7.4 \mu$; *Casmerodius*, $15.0 \mu \times 7.9 \mu$; *Tigrisoma*, $15.8 \mu \times 10.2 \mu$); falconids (*Falco*, $11.8 \mu \times 7.3 \mu$ compared with *Caracara*, $14.1 \mu \times 8.7 \mu$) and alcedinids (*Chloroceryle aenea*, $12.6 \mu \times 6.7 \mu$ compared with *Chloroceryle amazona*, $14.0 \mu \times 7.6 \mu$). The erythrocyte size is nearly the same in the three species of cathartids and in all parulids.

In most species the nucleus is more oblong than is the cytosome (compare the ratios). The cotingid nucleus is unusually elongated except in *Tityra*.

We can assume that, in order to permit free movement of erythrocytes, the diameter of the smallest capillaries must be no less than the smaller diameter of the erythrocytes. With this assumption in mind, it is interesting to compare the minimal diameters of different species. In our series this range is from 5.6μ in *Campylopterus* to 10.2μ in *Tigrisoma*, but in a majority of birds the range is much smaller, being 6μ to 8μ .

DISCUSSION

Cytosome size.—Since the erythrocyte is the most important carrier of oxygen and CO_2 , its surface area to size ratio is a determining factor in the exchange of these gases with the tissues. Thus, a small corpuscle offers the possibility of a greater rate of exchange than a larger one. Likewise an elliptical body is more efficient than a spherical one of the same volume. Avian erythrocytes are efficient in both of these respects. The high rate of metabolism of birds demands greater efficiency of gaseous exchange than that of many vertebrates. The difference in metabolic rate among birds is partially reflected by erythrocyte size, smaller species tending to possess smaller erythrocytes than do larger species.

It is interesting, however, that some of the largest birds, the Struthionidae, have erythrocytes no larger than those of the species reported by us. Gulliver (1840) gave the following values: Ostrich (*Struthio camelus*) cytosome— $15.4 \mu \times 8.5 \mu$, nucleus— $7.9 \mu \times 2.8 \mu$; Emu (*Dromiceius n. hollandiae*) cytosome— $15.0 \mu \times 8.4 \mu$; *Rhea americana*, cytosome— $13.4 \mu \times 7.8 \mu$. But later (1846), for the Cassowary (*Casuaris*), he gave the size of the erythrocyte as $17.5 \mu \times 9.1 \mu$.

In general our findings agree with those of Cleland and Johnston

(1912), that the lower forms possess the larger erythrocytes. Many of their families are the same as ours but the species are not.

Bartsch *et al.* (1937) show values in their tables little different from ours. However, in their text they give a much greater range than those in their tables.

Comparison of the erythrocytes of birds with other warm-blooded animals is interesting. In mammals their diameters range from 5 μ and 6 μ (horse, cow, pig, mouse, and rat), to 7 μ and 7.4 μ (chimpanzee, woodchuck, and llama) (Wintrobe, 1961).

The red cell count in mammals is high, ranging from 6,300,000 (chimpanzee) to 15,000,000 per cubic mm (llama), while in birds it is 2,810,000 (fowl) to 3,530,000 (pigeon) (Wintrobe, 1961).

Comparison of birds with their ancestral relatives, the reptiles, shows how great the divergence is between these groups. According to Wintrobe (1961), reptilian erythrocytes range from 18.1 $\mu \times 8.7 \mu$ (turtle) to 23.2 $\mu \times 12.1 \mu$ (alligator) while the red cell count is 740,000 (turtle) to 670,000 (alligator) per cubic mm. These differences indicate how far ahead birds are on the metabolic scale.

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SUMMARY

Measurements of erythrocytes and their nuclei were made in 124 species of birds in 46 families collected in Panamá and the United States. The range of the erythrocytes extended from 10.7 $\mu \times 6.1 \mu$ in *Selasphorus* to 15.8 $\mu \times 10.2 \mu$ in *Tigrisoma*. The lower forms have the largest erythrocytes. In most species the nucleus is more oblong than is the cytosome. In a few families the erythrocytes are smaller in the smaller species. The cytosomes of trochilids and passerine birds, in keeping with their high rate of metabolism, are smaller than those in many other bird groups. Based on the assumption that the width of the erythrocytes indicates the minimum diameter of the capillaries, trochilids and some passerines must possess the smallest capillaries.

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