of bark. New holes and enlargements were made above the topmost row. Observations made within 12 feet with binoculars (7 × 50 Zeiss) showed that sapsuckers fed on sap which ran down from the leafy branch where it joined the main trunk. A bird might cling ½ hour before a well, alternately resting and feeding. These findings confirm those of Bolles (Auk, **8**: 256–270, 1891). In winter, sap stored in the trunk may be more stationary so that a pattern of rows well separated yields the most. Feeding sapsuckers, however, are forced to move about as there is no heavy flow at any one point. Above observations suggest that freezing nights and thawing days favor movement of sap. The volume and watery nature of the sapsucker's excreta may have resulted from an intake limited to sap. In summer feeding, solid food along with sap is afforded by occasional insects.—LAWRENCE KILHAM, *7815 Aberdeen Road, Bethesda, Md.*

Further Notes on Alcoholic Specimens.—In a recent issue of 'The Auk' (**72**: 300–303, 1955), I made suggestions for the preparation of alcoholic specimens and skeletons of birds. I have since received several inquiries regarding the shipment of spirit specimens. Collectors on expedition and foreign correspondents usually do not have large quantities of a preservative available. They also are concerned about the expense involved in shipping specimens in large fluid-filled containers. There is, as a matter of fact, a simple, inexpensive way to mail specimens. They do not need to be immersed in fluid while in transit. After the specimen has been in the fixative for three or more days (depending on the size of the specimen and the method of treatment), it may be wrapped in cotton saturated with the fixative, placed in a double-walled polyethylene-type bag, and mailed in a wooden or heavy cardboard box. Two plastic bags, one placed inside the other, serve this purpose admirably. Or, a tubular section of plastic material can easily be transformed into a double-walled sack by tying a cord around the tube at its midpoint and then doubling one half over the other. When the open ends are tied securely, the fumes from the saturated cotton will maintain the specimen in excellent condition for several months. I have received well-preserved specimens from Africa prepared in this manner. It is important that the bags be sealed to prevent seepage and excessive evaporation of the fixative. In order to prevent punctures of the plastic, sharp beaks and claws should be wrapped with cotton or cloth.

My previous note dealt with the preparation of spirit specimens for gross dissection. For this purpose, preservation in alcohol or embalming fluid is desirable because these fluids are more pleasant to work with than is formalin. Certain constituents of some embalming fluid formulae, however, may interfere seriously with microtechnic staining methods. If one intends to do any microscopic work, therefore, it is best to use another fixing agent. Experience indicates that one of the best fixatives for this sort of work is buffered 10 per cent formalin. For the convenience of ornithologists, I quote here the formula as given by Lillie (1954. Histopathologic technic and practical histochemistry. Blakiston Co., p. 34). "Neutral buffered formaldehyde solution (pH 7.0). 37–40 per cent formaldehyde solution, 100 cc.; water, 900 cc.; acid sodium phosphate, monohydrate, 4 grams; anhydrous disodium phosphate, 6.5 grams." After material has been removed for microscopic study, the specimen may be washed in running water to remove excess formalin in order to reduce skin irritation when handling in gross dissection.

Gale and Geary (Anat. Rec., **124**: 95–99, 1956) recently presented an embalming fluid formula, which they found gave satisfactory preservation for histologic diagnosis in all tissues, although there was autolysis of the mucosa of the gastro-intestinal tract.—ANDREW J. BERGER, *Department of Anatomy, University of Michigan Medical School, Ann Arbor, Michigan.*